

From: Fred Kibenge [Kibenge@upeu.ca]
Sent: November-23-11 12:38 PM
To: Jonathan Coady
Subject: Fwd: Re: Samples
Attachments: Alexandra Morton Samples (HERRING and
SOCKEYE)_VT10312011_OCTOBER31 2011.pdf

e-mail 26

>>> Fred Kibenge 11/3/2011 1:34 PM >>>
Hello Alexandra,

Please find attached the real-time RT-PCR results of your most recent
set of samples.

Fred.

>>> Alexandra Morton <gorbuscha@gmail.com> 31/10/2011 1:35 PM >>>
Dear Fred

Hope you have a restful weekend and are not buried in snow.

I see from the FedEx tracking that my most recent set of samples have
arrived at your lab.

Please let me know if you have got them and if you will be permitted to
test
them for ISA virus.

Patient

Alex

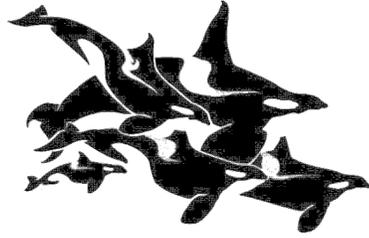
AVC Lab #	Sample ID	ISAV seg 8 Probe, Cts	
		Detects all ISAV	
VT 10312011-69	Sokeye smolts-Gill and Heart 1		0
VT 10312011-70	Sokeye smolts-Gill and Heart 3		0
VT 10312011-71	Sokeye smolts-Gill and Heart 4		0
VT 10312011-72	Sokeye smolts-Gill and Heart 5		0
VT 10312011-73	Sokeye smolts-Gill and Heart 6		0
VT 10312011-74	Herring - heart-17		0
VT 10312011-75	Herring - heart-18		0
VT 10312011-76	Herring - heart-19		0
VT 10312011-77	Herring - heart-20		0
VT 10312011-78	Herring - heart-21		0
ADL-ISAV (European genotype)			18.65
NBISAV01 (North American genotype)			17.95
NTC (water)			0

EXPLANATORY NOTES:

1. All samples were tested for ISAV using real-time RT-PCR with TaqMan probe for ISAV segment 8. The result is the number of PCR cycles to reach reliable detection of product (cycle threshold or Ct).
2. Based on this PCR test, all 10 tissue samples provided to the laboratory were RT-PCR negative. This test means none of the tissues contained ISAV sequences of genome segment 8.
3. The laboratory did not participate in the collection of the samples or in the custody of the samples prior to receipt of the samples. The laboratory therefore cannot guarantee the integrity of the samples.
4. For convenience, the samples are identified using the labels provided by the party who requested testing by the laboratory.
5. Case history on the submitted samples was requested and is attached to this report.
6. The samples were tested as received at the laboratory.
7. In accordance with the Health of Animals Act, the test results have been reported to representatives from the CFIA by the laboratory.

INTERPRETATION:

1. Ct up to 40 are positive. Ct between 40.1 and 45 are considered suspicious. Sample is negative if there is no Ct value.



Raincoast Research Society, Box 399, Sointula, BC V0N 3E0

**Samples shipped October 27, 2011 to:
Dr. Fred Kibenge, Atlantic Veterinary College, PEI.**

Owner name and address: Alexandra Morton, Box 399, Sointula, BC V0N 3E0

Vials # 17, 18, 19, 20, 21

Date collected: Oct. 26, 2011

Location: South side of Malcolm Island, Pacific Management Area 12

Species: 5 Herring

Storage: Fish were placed in RNALater less than one hour after death
Shipped the next day

Clinical symptoms/lesions: bleeding from fins, blood speckled eyes, rolling
over and sinking

Samples: hearts – whole, one per container

Vials # 1, 3, 4, 5, 6, (#2 omitted)

Date collected: June 2011

Location: Okisollo Channel, Pacific Management Area 13

Species: 5 Sockeye Smolts

Storage: Fish were placed in household type freezer and put in RNALater
October 26

Shipped the next day

Clinical symptoms/lesions: bleeding from fins, popped eyeballs

Samples: hearts and gills, one fish per container

Thank you,

Alexandra Morton
250-973-2306
250-974-7086 (cell)
gorbuscha@gmail.com