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Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: May 20, 2003 9:16 AM
To: JonesS@pac.dfo-mpo.gc.ca
Subject: ASK-2 cell line and viral RNA

Follow Up Flag: Follow up
Flag Status: Red

Hello Simon,

Today I shipped three 75 cm² flasks with ASK-2 cell monolayers.
The cells are 2 days old today.

To use, we split them 1:2 when the monolayer is confluent. Growth medium is L-15 + 1 mM L-glutamine + 10% FBS (from Sigma .. Hybridoma tested) + 1x antibiotics.
Maintenance medium is same with FBS reduced to 5%.

These cells grow very slowly but once confluent they are very susceptible to ISAV (I have seen CPE come up in 2 days!).
However, I do not know how good they are for primary virus isolation.

I have also include ISAV isolate NBISA01 RNA in formamide.
To use, precipitate with 2.5 x volume 100% ethanol in high salt and then resuspend in RNase-free water.

The shipment was sent Federal Express. The FedEx track number is as follows:
4967 8015 7201.

Let me know when you receive it.

Regards,

Fred.

Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: October 3, 2003 10:18 AM
To: JonesS@pac.dfo-mpo.gc.ca
Subject: RE: visit

Follow Up Flag: Follow up
Flag Status: Red

Hello Simon,

Good to hear from you. I would be happy to run those samples through our RT-PCR system here. We also use the segment 8 F5/R5 primers. We clone our products using the Invitrogen TOPO TA Cloning kit and use a commercial company for sequencing. Feel free to send the samples along and I will let you know what we find.

Best regards.

Fred.

(by the way, I plan to be in Nanaimo for the Thanksgiving weekend. Hopefully I will see you then?).

Date sent: Fri, 03 Oct 2003 10:08:55 -0700
From: JonesS@pac.dfo-mpo.gc.ca
Subject: RE: visit
To: kibenge@upei.ca

> Hi Fred:
> I hope all is going well. I am hoping you will be interested in
> collaborating with us on our virus surveillance studies. We have been
> screening Pacific salmon for viral pathogens and have some samples
> that we would like to have confirmed for ISA virus. We noticed a
> positive RTPCR reaction using the segment 8 (f5/r5) primers and were
> interested in having these observations confirmed, with sequencing of
> the reaction products. I have 20 samples that I would like to send you.
> Looking forward to your reply.
> Best wishes,
> Simon
>
> Dr. Simon R.M. Jones
> Department of Fisheries and Oceans
> Pacific Biological Station
> 3190 Hammond Bay Road
> Nanaimo, British Columbia
> V9T 6N7, Canada
>
> tel: 250 729 8351
> fax: 250 756 7053
> joness@pac.dfo-mpo.gc.ca
> http://www.pac.dfo-mpo.gc.ca/sci/aqua/profiles/jones_e.htm
>
>
>
>
>

> -----Original Message-----
> From: Dr. Fred Kibenge [mailto:kibenge@upei.ca]
> Sent: July 9, 2003 8:27 AM
> To: JonesS@pac.dfo-mpo.gc.ca
> Subject: Re: visit

>
>
> Hi Simon,
>
> Good to hear from you. I am still interested in presenting a seminar
> and further discussions on collaboration.
>
> The Seminar title is "Biochemistry, Aetiopathogenesis and Immunology
> of Infectious Salmon Anaemia Virus".
>
> All the best.
>
> Fred.

Hi Simon,
I just wanted to let you know that I received the 20 samples (1A-10A and 1B-10B) in good condition, and will soon start working on them.

All the best,
Fred.

From: Dr. Fred Kibenge (kibenge@upei.ca)
To: jones@pac.dfo-mpc.gc.ca
Subject: RE: Virus
Sent: Friday, October 1, 2004 11:17:45 AM
Date sent: Fri, 01 Oct 2004 10:08:32 -0700

Hi Simon,
>
> Good to hear from you. I would be happy to run those samples through
> our RT-PCR system here. We also use the segment 8
> RT-PCR primers. We clone our products using the Intronspan T700 TA
> Cloning kit and use a commercial company for sequencing.
> Feel free to send the samples along and I will let you know what we
> find.

Best regards,

Fred.

> (by the way, I plan to be in Moncton for the Thanksgiving weekend.
> Hopefully I will see you then!).

Date sent: Fri, 01 Oct 2004 10:08:32 -0700
From: jones@pac.dfo-mpc.gc.ca
Subject: RE: Virus
To: kibenge@upei.ca

Hi Fred:
> I hope all is going well. I am hoping you will be interested in
> collaborating with us on our virus surveillance studies. We have
> been screening Pacific salmon for viral pathogens and have some
> samples that we would like to have confirmed for ISA virus. We
> noticed a positive RT-PCR reaction using the segment 8 (F5/F7)
> primers and were interested in having these observations confirmed,
> with sequencing of the reaction products. I have 20 samples that I would like to
send you.
> Looking forward to your reply.

Best wishes,
Simon

Dr. Simon M. Jones
> Department of Fisheries and Oceans
> Pacific Biological Station
> 3190 Hammond Bay Road

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Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: October 7, 2003 6:36 AM
To: JonesS@pac.dfo-mpo.gc.ca
Subject: RE: visit

Follow Up Flag: Follow up
Flag Status: Red

Hello Simon,

I just wanted to let you know that I received the 20 samples (1A-10A and 1B-10B) in good condition, and will soon start working on them.

All the best.

Fred.

From: Dr. Fred Kibenge <kibenge@ACAD1.CS.UPEI.CA>
To: JonesS@pac.dfo-mpo.gc.ca
Subject: RE: visit
Send reply to: kibenge@upei.ca
Date sent: Fri, 3 Oct 2003 17:17:48 ADT

> Hello Simon,

>

> Good to hear from you. I would be happy to run those samples through
> our RT-PCR system here. We also use the segment 8
> F5/R5 primers. We clone our products using the Invitrogen TOPO TA
> Cloning kit and use a commercial company for sequencing.
> Feel free to send the samples along and I will let you know what we
> find.

>

> Best regards.

>

> Fred.

>

> (by the way, I plan to be in Nanaimo for the Thanksgiving weekend.
> Hopefully I will see you then?).

>

> Date sent: Fri, 03 Oct 2003 10:08:55 -0700
> From: JonesS@pac.dfo-mpo.gc.ca
> Subject: RE: visit
> To: kibenge@upei.ca

>

> > Hi Fred:

> > I hope all is going well. I am hoping you will be interested in
> > collaborating with us on our virus surveillance studies. We have
> > been screening Pacific salmon for viral pathogens and have some
> > samples that we would like to have confirmed for ISA virus. We
> > noticed a positive RTPCR reaction using the segment 8 (f5/r5)
> > primers and were interested in having these observations confirmed,
> > with sequencing of the reaction products. I have 20 samples that I would like to
> send you.

> > Looking forward to your reply.

> > Best wishes,

> > Simon

> >

> > Dr. Simon R.M. Jones
> > Department of Fisheries and Oceans
> > Pacific Biological Station
> > 3190 Hammond Bay Road

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> > Nanaimo, British Columbia
> > V9T 6N7, Canada

> > tel: 250 729 8351
> > fax: 250 756 7053
> > jones@pac.dfo-mpo.gc.ca
> > http://www.pac.dfo-mpo.gc.ca/sci/aqua/profiles/jones_e.htm

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> >
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> > -----Original Message-----

> > From: Dr. Fred Kibenge [mailto:kibenge@upei.ca]
> > Sent: July 9, 2003 8:27 AM
> > To: Jones@pac.dfo-mpo.gc.ca
> > Subject: Re: visit

> >
> >
> >

> > Hi Simon,

> > Good to hear from you. I am still interested in presenting a
> > seminar and further discussions on collaboration.

> > The Seminar title is "Biochemistry, Aetiopathogenesis and Immunology
> > of Infectious Salmon Anaemia Virus".

> > All the best.

> > Fred.

>
>

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Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: October 21, 2003 1:57 AM
To: JonesS@pac.dfo-mpo.gc.ca
Cc: KibengeM@pac.dfo-mpo.gc.ca
Subject: gels

Follow Up Flag: Follow up
Flag Status: Red

Hello Simon,

Attached are some gels of the testing on the 20 samples.
My pictures are clearer than this (clear enough to cut the bands out) but I thought you might want to have an idea of the intensity of the bands compared to the positive controls?

All the best.

Fred.

Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: October 21, 2003 1:57 AM
To: JonesS@pac.dfo-mpo.gc.ca
Cc: KibengeM@pac.dfo-mpo.gc.ca
Subject: gels

Follow Up Flag: Follow up
Flag Status: Red

Attachments: DFO gels.doc



DFO gels.doc (22 KB)

- * This message contains the file 'DFO gels.doc', which has been
- * uuencoded. If you are using Pegasus Mail, then you can use
- * the browser's eXtract function to lift the original contents
- * out to a file, otherwise you will have to extract the message
- * and uudecode it manually.

High Sea Chinnok Salmon samples checked for ISAV using RT-PCR (samples received from DFO, PBS, Dr. Simon Jones/Dr. Molly Kibenge).

Figure 1: Segment 8 primers (F5/R5). All positive samples ran on one gel.

Lanes:

1. 1kb ladder (Invitrogen)
2. Sample 3B
3. Sample 4B
4. Sample 10B
5. NBISA01 +ve control
6. Sample 1A
7. Sample 2A
8. Sample 3B
9. Sample 4B
10. NBISA01 +ve control

Figure 2: Segment 7 ORF1 primers

Lanes:

1. 1kb ladder (Invitrogen)
2. Sample 1A -ve
3. Sample 2A +ve (400bp)
4. Sample 3B -ve
5. Sample 4B +ve (400bp)
6. Sample 5B -ve
7. Sample 6B +ve (400bp)
8. Sample 10B +ve (400bp)
9. NBISA01 +ve control (903bp)

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Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: October 21, 2003 1:57 AM
To: JonesS@pac.dfo-mpo.gc.ca
Cc: KibengeM@pac.dfo-mpo.gc.ca
Subject: gels

Follow Up Flag: Follow up
Flag Status: Red

Attachments: DFO 100703 gels.ppt



DFO 100703
gels.ppt (140 KB)

* This message contains the file 'DFO 100703 gels.ppt', which has been
* uuencoded. If you are using Pegasus Mail, then you can use
* the browser's eXtract function to lift the original contents
* out to a file, otherwise you will have to extract the message
* and uudecode it manually.

High Sea Chinnok Salmon samples checked for ISAV using RT-PCR (samples received from DFO, PBS, Dr. Simon Jones/Dr. Molly Kibenge).

Figure 1: Segment 8 primers (F5/R5). All +ve samples ran on one gel.

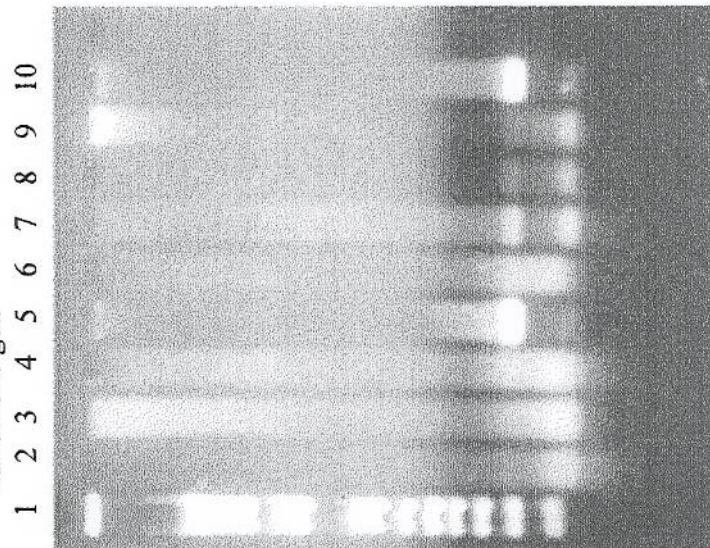
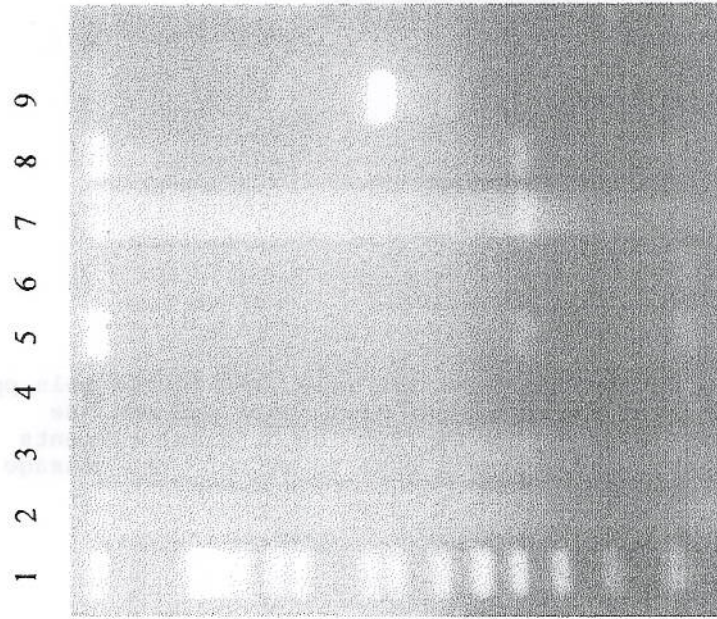


Figure 2: Segment 7 ORF1 primers



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Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: November 5, 2003 1:34 PM
To: JonesS@pac.dfo-mpo.gc.ca
Cc: KibengeM@pac.dfo-mpo.gc.ca
Subject: sequence analysis of segment 7 products

Follow Up Flag: Follow up
Flag Status: Red

Hello Simon,

* Attached is the information I have obtained on the segment 7 clones. You will note that I am being very cautious.

All the best.

Fred.

Sample ID	Segment 7 product (bp)	Size (bp)
1A	+	~400bp
2A	+	~400bp
3A	+	~400bp
4A	+	~400bp
5A	+	~400bp
6A	+	~400bp
7A	+	~400bp
8A	+	~400bp
9A	+	~400bp
10A	+	~400bp
1B	+	~400bp
2B	+	~400bp
3B	+	~400bp
4B	+	~400bp
5B	+	~400bp
6B	+	~400bp
7B	+	~400bp
8B	+	~400bp
9B	+	~400bp
10B	+	~400bp
MB12A01 +ve control	+	~400bp

(12)

*

High Sea Chinnok Salmon samples labeled 1A to 10A and 1B to 10B, checked for ISAV using RT-PCR (samples received from DFO, PBS, Dr. Simon Jones/Dr. Molly Kibenge).

Sample ID	Segment 8 primers (F5/R5)	Segment 7 ORF1 primers
1A	+ ve (~220bp)	- ve
2A	+ ve (~220bp)	+ ve (~400bp)
3A	weak + ve (~220bp)	not done
4A	- ve	not done
5A	- ve	not done
6A	- ve	not done
7A	- ve	not done
8A	- ve	not done
9A	- ve	not done
10A	- ve	not done
1B	- ve	not done
2B	- ve	not done
3B	+ ve (~220bp)	- ve
4B	+ ve (~220bp)	+ ve (~400bp)
5B	- ve	- ve
6B	- ve	+ ve (~400bp)
7B	- ve	not done
8B	- ve	not done
9B	- ve	not done
10B	+ ve (~220bp)	+ ve (~400bp)
NBISA01 +ve control	+ ve (~220bp)	+ ve (903bp)

- ve denotes no PCR product seen; + ve denotes PCR product (size in base pairs) seen.

Sequence Analysis of the RT-PCR products of the Segment 7 ORF1 primers:

All 5 RT-PCR products obtained were subcloned using the TOPO TA cloning kit. The DNA inserts were then sequenced.

The NBISA01 +ve control DNA insert was 903 bp long.

All inserts of the DFO samples were 377 bp long.

The DFO samples were 99.7% identical and corresponded to the ISAV Segment 7 ORF2 product. They had a 95.8% identity with the NBISA01 +ve control, and 99.7% identity with ISAV strain 810/9/99 from Norway. The sequences are given on page 3, and sequence comparisons using the FASTA program are attached in the next 6 pages (pages 4-9).

[These particular DFO samples were processed separately by my self together with ISAV isolate NBISA01 as positive control. So, I can rule out cross contamination during RNA extraction. However, I also have a graduate student working with both New Brunswick and European (Norway, Scotland, Nova Scotia) ISAV isolates, and cannot conclusively rule out the possibility (however small!) of reagent contamination. Reagent contamination is unlikely since the RT-PCR product was not in all samples tested. For this reason, I am sending you the sequence of the Segment 7 ORF1 primers that I used so that you can try them on your samples. Also, I wish to encourage you to send other blind samples to me for RT-PCR testing so that you can confidently interpret your results].

Segment 7 ORF1 primers:

For the purpose of evaluating individual ORFs, we separated the otherwise partial overlapping ORFs of segment 7 based on the gene expression model suggested by Ritchie *et al.* (2002), of partial splicing of ORF1 mRNA and a frameshift such that ORFs 1 and 2 products share the first 22 amino acid residues.

ISAV SEG7 (ORF1) FOR (23 mer) 5'-ATGGATTTCACCAAAGTGTATGG-3'	1-23,	AF328627
ISAV SEG7 (ORF1) REV (23 mer) 5'-TCACATTCTGAAGTGAAGTCCAG-3'	881-903,	AF328627

ISAV SEG7 (ORF2) FOR (82 mer) 5'-ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAA		
AACTTCACGAAAAGACAAGGTGGCTTCTTCCTGTCGG-3'	1-63/590-608,	AF328627

Ritchie, R. J., A. Bardiot, K. Melville, S. Griffiths, C. O. Cunningham, and M. Snow. 2002. Identification and characterization of the genomic segment 7 of the infectious salmon anaemia virus genome. *Virus Res.* **84**:161-70.

(E1) 14

>2A DFO Seg 7 ORF1 reverse (2A-2)

ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAACCTTCACGGAAAA
GACAAGGTGGCTTCTTTCCTGTCGGGGCTCAAAGGTTCTTGGGGAGGATGGTATCTCA
AGTACGTCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAAC
GATTCTATGACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAA
ATGGACAGAGACGGCGTATCATTATCTACGAGAAGCCTAGCATCTACCATAGTGAT
GGGTGCACTGGGACAGCATCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGG
AGTTGAGCTTAGGGCTGGACTTCACTTCAGAATGTGA

>4B DFO Seg 7 ORF1 (4B-1)

ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAACCTTCACGGAAAA
GACAAGGTGGCTTCTTTCCTGTCGGGGCTCAAAGGTTCTTGGGGAGGATGGTATCTCA
AGTACGTCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAAC
GATTCTATGACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAA
ATGGACAGAGACGGCGTATCATTATCTACGAGAAGCCTAGCATCTACCATAGTGAT
GGGTGCACTGGGACAGCATCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGG
AGTTGAGCTTAGGGCTGGACTTCACTTCAGAATGTGA

>6B DFO Seg 7 ORF1 (6B-2)

ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAACCTTCACGGAAAA
GACAAGGTGGCTTCTTTCCTGTCGGGGCTCAAAGGTTCTTGGGGAGGATGGTATCTCA
AGTACGTCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAAC
GATTCTATGACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAA
ATGGACAGAGACGGCGTATCATTATCTACGAGAAGCCTAGCATCTACCATAGTGAT
GGGTGCACTGGGACAGCATCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGG
AGTTGAGCTTAGGGCTGGACTTCACTTCAGAATGTGA

>10B DFO Seg 7 ORF1 (10B-1)

ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAACCTTCACGGAAAA
GACAAGGTGGCTTCTTTCCTGTCGGGGCTCAAAGGTTCTTGGGGAGGATGGTATCTCA
AGTACGTCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAAC
GATTCTATGACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAA
ATGGACAGAGACGGCGTATCATTATCTACGAGAAGCCTAGCATCTACCATAGTGAT
GGGTGCACTGGGACAGCAGCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTG
GAGTTGAGCTTAGGGCTGGACTTCACTTCAGAATGTGA

A:DFO4B.SEQ, 377 nt vs A:DFO10B.SEQ library
using DNA matrix

377 residues in 1 sequences
1 scores better than 59 saved, ktup: 4
The best scores are:

initn	init1	opt
1501	1501	1501
1501	1501	1501

>10B DFO Seg 7 ORF1 (10B-1)
99.7% identity in 377 nt overlap

	10	20	30	40	50	60
4B DFO	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TTACGGAAGAC				
X:
10B	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TTACGGAAGAC				
	10	20	30	40	50	60
	70	80	90	100	110	120
4B DFO	AAGGTGGCTTCTTTCCTGTGCGGGCTCAAAGGTTCTTGGGGAGGATGGTATCTCAAGTACG					
:
10B	AAGGTGGCTTCTTTCCTGTGCGGGCTCAAAGGTTCTTGGGGAGGATGGTATCTCAAGTACG					
	70	80	90	100	110	120
	130	140	150	160	170	180
4B DFO	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAACGATTCTATG					
:
10B	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAACGATTCTATG					
	130	140	150	160	170	180
	190	200	210	220	230	240
4B DFO	ACAGAGCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
:
10B	ACAGAGCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
	190	200	210	220	230	240
	250	260	270	280	290	300
4B DFO	GCGTATCATTCATCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG					
:
10B	GCGTATCATTCATCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG					
	250	260	270	280	290	300
	310	320	330	340	350	360
4B DFO	CATCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
:
10B	CAGCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
	310	320	330	340	350	360
	370					
4B DFO	TTCACTTCAGAATGTGA					
:X					
10B	TTCACTTCAGAATGTGA					
	370					

Library scan: 0:00:00 total CPU time: 0:00:19

(12)

(16) X

A:DFO6B.SEQ, 377 nt vs A:DFO10B.SEQ library
using DNA matrix

377 residues in 1 sequences
1 scores better than 59 saved, ktup: 4
The best scores are: initn initl opt
>10B DFO Seg 7 ORF1 (10B-1) 1501 1501 1501

>10B DFO Seg 7 ORF1 (10B-1) 1501 1501 1501
99.7% identity in 377 nt overlap

	10	20	30	40	50	60
6B DFO	ATGGATTTACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TTACCGGAAAAGAC				
	X::					
10B	ATGGATTTACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TTACCGGAAAAGAC				
	10	20	30	40	50	60
	70	80	90	100	110	120
6B DFO	AAGGTGGCTTCTTTCTGTCGGGCTCAAAGGTTCTGGGGAGGATGGTATCTCAAGTACG					
	::					
10B	AAGGTGGCTTCTTTCTGTCGGGCTCAAAGGTTCTGGGGAGGATGGTATCTCAAGTACG					
	70	80	90	100	110	120
	130	140	150	160	170	180
6B DFO	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTCAATTGTCAATCAACGATTCTATG					
	::					
10B	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTCAATTGTCAATCAACGATTCTATG					
	130	140	150	160	170	180
	190	200	210	220	230	240
6B DFO	ACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
	::					
10B	ACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
	190	200	210	220	230	240
	250	260	270	280	290	300
6B DFO	GCGTATCATTCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG					
	::					
10B	GCGTATCATTCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG					
	250	260	270	280	290	300
	310	320	330	340	350	360
6B DFO	CATCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
	:: ::					
10B	CAGCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
	310	320	330	340	350	360
	370					
6B DFO	TTCACTTCAGAATGTGA					
	::::::::::::::::::::X					
10B	TTCACTTCAGAATGTGA					
	370					

Library scan: 0:00:00 total CPU time: 0:01:46

(17)

a:dfol0b.seq, 377 nt vs a:dfo2ar.seq library
using DNA matrix

377 residues in 1 sequences
1 scores better than 59 saved, ktup: 4
The best scores are: initn initl opt
>2A DFO reverse 1501 1501 1501
 1501 1501 1501
>2A DFO reverse
99.7% identity in 377 nt overlap

	10	20	30	40	50	60
10B DF	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAACTTCACGGAAAAGAC					
	X:.....					
2A	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAACTTCACGGAAAAGAC					
	10 20 30 40 50 60					
	70 80 90 100 110 120					
10B DF	AAGGTGGCTTCTTTCCTGTGCGGGCTCAAAGGTTCTGGGGAGGATGGTATCTCAAGTACG					
					
2A	AAGGTGGCTTCTTTCCTGTGCGGGCTCAAAGGTTCTGGGGAGGATGGTATCTCAAGTACG					
	70 80 90 100 110 120					
	130 140 150 160 170 180					
10B DF	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAACGATTCTATG					
					
2A	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAACGATTCTATG					
	130 140 150 160 170 180					
	190 200 210 220 230 240					
10B DF	ACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
					
2A	ACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
	190 200 210 220 230 240					
	250 260 270 280 290 300					
10B DF	GCGTATCATTTCATCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG					
					
2A	GCGTATCATTTCATCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG					
	250 260 270 280 290 300					
	310 320 330 340 350 360					
10B DF	CAGCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
	::					
2A	CATCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
	310 320 330 340 350 360					
	370					
10B DF	TTCACCTCAGAATGTGA					
X					
2A	TTCACCTCAGAATGTGA					
	370					

Library scan: 0:00:00 total CPU time: 0:00:16

A:DF010B.SEQ, 377 nt vs A:810M2.SEQ library
using DNA matrix

485 residues in 1 sequences
1 scores better than 59 saved, ktup: 4
The best scores are: initn init1 opt
>810 SEG. 7 ORF2 (SE5058) 1501 1501 1501

>810 SEG. 7 ORF2 (SE5058) 1501 1501 1501
99.7% identity in 377 nt overlap

		10	20	30	40	50	60
10B DF	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TT	CACGGAAAAGAC				
	X	:	:	:	:	:	:
810	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TT	CACGGAAAAGAC				
		10	20	30	40	50	60
		70	80	90	100	110	120
10B DF	AAGGTGGCTTCTTTCCTGTCGGGCTCAAAGGTTCTGGGGAGGATGGTATCTCAAGTACG						
	:	:	:	:	:	:	:
810	AAGGTGGCTTCTTTCCTGTCGGGCTCAAAGGTTCTGGGGAGGATGGTATCTCAAGTACG						
		70	80	90	100	110	120
		130	140	150	160	170	180
10B DF	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTCAATTGTCAATCAACGATTCTATG						
	:	:	:	:	:	:	:
810	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTCAATTGTCAATCAACGATTCTATG						
		130	140	150	160	170	180
		190	200	210	220	230	240
10B DF	ACAGAGCCCAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG						
	:	:	:	:	:	:	:
810	ACAGAGCCCAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG						
		190	200	210	220	230	240
		250	260	270	280	290	300
10B DF	GCGTATCATTCATCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG						
	:	:	:	:	:	:	:
810	GCGTATCATTCATCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG						
		250	260	270	280	290	300
		310	320	330	340	350	360
10B DF	CAGCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC						
	:	:	:	:	:	:	:
810	CATCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC						
		310	320	330	340	350	360
		370					
10B DF	TTCACCTCAGAATGTGA						
	:	:	:	:	:	:	:
810	TTCACCTCAGAATGTGATTGGCTGAAAACATGTTTTGTAAACAAGAATTTTGTGTTTTTG						
		370	380	390	400	410	420

Library scan: 0:00:00 total CPU time: 0:00:37

a:dfol0b.seq, 377 nt vs a:nbisadfo.seq library
using DNA matrix

903 residues in 1 sequences
1 scores better than 59 saved, ktup: 4
The best scores are:
>NBISA-1 (SF2912; DFO Seg 7 ORF1 NBISA-1)
>NBISA-1 (SF2912; DFO Seg 7 ORF1 NBISA-1)
94.9% identity in 316 nt overlap

initn initl opt
1152 1152 1152
1152 1152 1152

	40	50	60	70	80	90
10B DF	TTGACCAACTAAAACTTCACGGAAAAGACAAGGTGGCTTCTTTTCCTGTCGGGGCTCAAAGG					
	X::::::::::::::::::::::::::::					
NBISA-	TTTCCTTGATGAACCTTGCTACTGTTGTTACAGGTGGCTTCTTTTCCTGTCGGACTCAAAGG					
	560	570	580	590	600	610
	100	110	120	130	140	150
10B DF	TTCCTGGGGAGGATGGTATCTCAAGTACGTCAGGTATGCTGGACCTCTTGAAGGATCAAG					
	::					
NBISA-	TTCCTGGGGAGGATGGTACCTCAAGTACGTCAGGTATGCTGGACCTCTTGCGGGATCAAG					
	620	630	640	650	660	670
	160	170	180	190	200	210
10B DF	TGGGTTCATTGTCAATCAACGATTCTATGACAGAGCCCAAAACAGAGCTGGATCCAGGGT					
	::					
NBISA-	TGGATTTCATTGTCAATCAACGATTCTACGACAGAGCCCAAAACAAGACTGGATCCAGGGT					
	680	690	700	710	720	730
	220	230	240	250	260	270
10B DF	TGTATCCATGGTTGAAATGGACAGAGACGGCGTATCATTCATCTACGAGAAGCCTAGCAT					
	::					
NBISA-	TGTATCCATGGTTGAAATGGACGGGACAGCGGCTTATCGTTCATCTACGAGAAGCCTAGCGT					
	740	750	760	770	780	790
	280	290	300	310	320	330
10B DF	CTACCATAGTGATGGGTGCACTGGGACAGCAGCGAGGGTCTGGAGACGGGATCACAATGA					
	::					
NBISA-	CTACCATAGTGATGGGTGCACTGGGTCAGCAGCGAGGTTCTGGAGACGGGATCGCAATGA					
	800	810	820	830	840	850
	340	350	360	370		
10B DF	GAGAGCTGGAGTTGAGCTTAGGGCTGGACTTCACTTCAGAATGTGA					
	::X					
NBISA-	GAGAGCTGGAGTTGAGCTTAGGGCTGGACTTCACTTCAGAATGTGA					
	860	870	880	890	900	

Library scan: 0:00:00 total CPU time: 0:00:28

20

X

A:DF010B.SEQ, 377 nt vs A:NBISAM2.SEQ library
using DNA matrix

485 residues in 1 sequences
1 scores better than 59 saved, ktup: 4
The best scores are: initn initl opt
>NBISA01 SEG. 7 ORF2 (SE5060) 1396 1396 1396
>NBISA01 SEG. 7 ORF2 (SE5060) 1396 1396 1396
95.8% identity in 377 nt overlap

	10	20	30	40	50	60
10B DF	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TTACGGAAAAGAC				
X:	:	:	:	:	:	:
NBISA0	ATGGATTTCACCAAAGTGTATGGTGTGCTGGTTGACCAACTAAAAC	TTACGGAAACAGAC				
	10	20	30	40	50	60
	70	80	90	100	110	120
10B DF	AAGGTGGCTTCTTTCCCTGTCGGGCTCAAAGGTTCCCTGGGGAGGATGGTATCTCAAGTACG					
:	:	:	:	:	:	:
NBISA0	AAGGTGGCTTCTTTCCCTGTCGGGCTCAAAGGTTCCCTGGGGAGGATGGTATCTCAAGTACG					
	70	80	90	100	110	120
	130	140	150	160	170	180
10B DF	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAACGATTCTATG					
:	:	:	:	:	:	:
NBISA0	TCAGGTATGCTGGACCTCTTGAAGGATCAAGTGGGTTTCATTGTCAATCAACGATTCTATG					
	130	140	150	160	170	180
	190	200	210	220	230	240
10B DF	ACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
:	:	:	:	:	:	:
NBISA0	ACAGAGCCCCAAAACAGAGCTGGATCCAGGGTTGTATCCATGGTTGAAATGGACAGAGACG					
	190	200	210	220	230	240
	250	260	270	280	290	300
10B DF	GCGTATCATTTCATCTACGAGAAGCCTAGCATCTACCATAGTGATGGGTGCACTGGGACAG					
:	:	:	:	:	:	:
NBISA0	GCTTATCGTTTCATCTACGAGAAGCCTAGCGTCTACCATAGTGATGGGTGCACTGGGTCAG					
	250	260	270	280	290	300
	310	320	330	340	350	360
10B DF	CAGCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
:	:	:	:	:	:	:
NBISA0	CAGCGAGGGTCTGGAGACGGGATCACAATGAGAGAGCTGGAGTTGAGCTTAGGGCTGGAC					
	310	320	330	340	350	360
	370					
10B DF	TTCACTTCAGAATGTGA					
:	:	:	:	:	:	:
NBISA0	TTCACTTCAGAATGTGATTGGTTGAAAACCTTGTTATGTAAACAAGAATTTTGTGTTTTG					
	370	380	390	400	410	420

Library scan: 0:00:00 total CPU time: 0:00:22

Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: November 10, 2003 8:41 AM
To: JonesS@pac.dfo-mpo.gc.ca
Cc: KibengeM@pac.dfo-mpo.gc.ca
Subject: sequence analysis of segment 8 products

Follow Up Flag: Follow up
Flag Status: Red

Hello Simon,

* Attached is the information I have obtained on the segment 8 clones. I suppose this result rules out the possibility of "reagent contamination" with lab. virus strains!

All the best.

Fred.

15

22

Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: November 10, 2003 8:41 AM
To: JonesS@pac.dfo-mpo.gc.ca
Cc: KibengeM@pac.dfo-mpo.gc.ca
Subject: sequence analysis of segment 8 products

Follow Up Flag: Follow up
Flag Status: Red

Attachments: DFO 100703 results.doc



DFO 100703
results.doc (108 KB..)

- * This message contains the file 'DFO 100703 results.doc', which has been
- * uuencoded. If you are using Pegasus Mail, then you can use
 - * the browser's eXtract function to lift the original contents
 - * out to a file, otherwise you will have to extract the message
 - * and uudecode it manually.

High Sea Chinnok Salmon samples labeled 1A to 10A and 1B to 10B, checked for ISAV using RT-PCR (samples received from DFO, PBS, Dr. Simon Jones/Dr. Molly Kibenge).

Sample ID	Segment 8 primers (F5/R5)	Segment 7 ORF1 primers
1A	+ ve (~220bp)	- ve
2A	+ ve (~220bp)	+ ve (~400bp)
3A	weak + ve (~220bp)	not done
4A	- ve	not done
5A	- ve	not done
6A	- ve	not done
7A	- ve	not done
8A	- ve	not done
9A	- ve	not done
10A	- ve	not done
1B	- ve	not done
2B	- ve	not done
3B	+ ve (~220bp)	- ve
4B	+ ve (~220bp)	+ ve (~400bp)
5B	- ve	- ve
6B	- ve	+ ve (~400bp)
7B	- ve	not done
8B	- ve	not done
9B	- ve	not done
10B	+ ve (~220bp)	+ ve (~400bp)
NBISA01 +ve control	+ ve (~220bp)	+ ve (903bp)

- ve denotes no PCR product seen; + ve denotes PCR product (size in base pairs) seen.

Sequence Analysis of the RT-PCR products of the Segment 8 F5/R5 primers:

Attempts were made to clone some of the RT-PCR products obtained using the TOPO TA cloning kit. The DNA inserts were then sequenced.

The NBISA01 +ve control DNA insert was 211 bp long.

Inserts of the DFO samples were variable in size and ranged from 116 bp long in clone 3B to 211 bp long in clone 2A. Of these clones, the most representative (based on sequence analysis) was clone 10B with a DNA insert of 187bp (since as shown in the alignment on page 13, clone 10B lines up very well with clone 2A; both 3B and 4B are missing internal sequences between the F5 and R5 primers because of an internal mis-priming or duplicate sequence; in clone 4B the F5 primer sequence is also duplicated, in tandem).

The DNA insert in clone 2A was the most similar to the NBISA01 +ve control DNA insert with 94.3% sequence identity. The other DFO samples sequenced (3B, 4B, and 10B) were only similar to the NBISA01 +ve control DNA insert in the primer regions, whereas they had considerable areas of sequence identity among themselves: 100% identity in 72 nucleotide overlap between 3B and 4B; 97.9% identity in 97 nucleotide overlap between 3B and 10B; and 95.9% identity in 97 nucleotide overlap between 4B and 10B. **Thus the "DFO B samples" would seem to contain a unique sequence, represented as clone 10B with a DNA insert of 187bp, amplified using the ISAV segment 8 F5/R5 primers. The smaller RT-PCR product of DFO 10B compared to ISAV NBISA01 is due to internal deletions in DFO 10B. I suppose this result rules out the possibility of "reagent contamination" with laboratory virus strains! For specific amplification of the virus in your samples, I suggest that you design new primers based on the DFO 10B clone sequence but excluding the F5 and R5 sequences. It would be interesting to see if these new primers amplify the original ISAV.**

The sequences are given on page 11, the sequence alignment of the 4 sample is shown on page 12, and the sequence comparisons using the FASTA program are given on pages 13-15.

This completes the analysis of the 20 High Sea Chinnok salmon samples. I look forward to more blind samples from you.

All the best.

Fred.

>2A DFO F5/R5 2A-#1 10/17 (SF3322)

GAAGAGTCAGGATGCCAAGACGCGGATGGTGGAGAGGAAAAGTGGGCAATGGTGT
ATGGTATGATTTACCAAGACATGGCGGAGGAGAAGACGATGTTGAAGGACCTGAAG
ACAATGCTACACAGCAGGATGCAGATGTATGCTCTAGGAGCGAGTTCGAAAGCCCT
GGAAACTTTAGAAAAGGCCATCGTCGCTGCAGATCATCGACTTC

>3B DFO F5/R5 3B-#2 10/9 (SF3323)

GAAGAGTCAGGATGCCAAGACGGAAGTCGCTGCAGATCATCGACTTCACTGAAGAG
TCAGGATGCCAAGACGGAAGTCGCTGCAGATCATCGACTTCGCTGCAGATCATCGA
CTTC

>4B DFO F5/R5 4B-#1 10/9 (SF3324)

GAAGAGTCAGGGATGCCAAGACG
GAAGAGTCAGGATGCCAAGACGAAGAGTCAGGATGCCAAGACGGAAGTCGCTGCA
GATCATCGACTTCACTGAAGAGTCAGGATGCCAAGACGCTCTCTCTCTCTCCCTCT
CTCTCTCTCTCTCTCTCTCTCGCTGCAGATCATCGACTTC

>10B DFO F5/R5 10B-#2 10/9 (SF3325)

GAATAGTCAGGATGCCAAGACGGAAGAGTCAGGATGCCAAGACGACTCGCTGCAGA
TCATCGACTTCACTGAAGAGTCAGGATGCCAAGACGGAAGAGTCAGGATCAAGACG
GAAGTCGCTGCAGATCATCGACTTCACTGAAGAGTCAGGATGCCAAGACGGAAGTC
GCTGCAGATCATCGACTTC

Page 1.1

1	15	16	30	31	45	46	60	61	75	76	90
1 3B	-----	-----	-----	-----	-----	-----	GAAGATCAGGATG	CCAAGACGGAAGTCG	CTGCAGATCATCGAC		44
2 10B	-----	-----	-----	-----	-----	-----	GAAGATCAGGATG	CCAAGACG--ACTCG	CTGCAGATCATCGAC		64
3 4B	GAAGAGTCAGGGATG	CCAAGACGGAAGAGT	CAGGATGCCAAGAC-	CAGGATGCCAAGAC-	CAGGATGCCAAGAC-	CAGGATGCCAAGAC-	CGGATGGTGGAGAGG	AAAAGTGGGCAATGG	-TGTAIGGTAT-GAT		88
4 2A	-----	-----	-----	-----	-----	-----	-----	-----	-----		65

Page 2.1

91	105	106	120	121	135	136	150	151	165	166	180
1 3B	TTCACTGAAGAGTCA	GGATGCCAAGACGGA	AG-----	TCGCTGCA	G----	ATCATCGACT-	-----	-----	TCGC	TGCAG-ATCATCGAC	113
2 10B	TTCACTGAAGAGTCA	GGATGCCAAGACGGA	AG-----	AGTCAGGA	TC--	AAGACGGAAG-	-----	-----	TCGC	TGCAG-ATCATCGAC	134
3 4B	TTCACTGAAGAGTCA	GGATGCCAAGACGCT	CTCTCTCTCTCTCC	TCT-CTCTCTCTCTC	TCT-CTCTCTCTCTC	TCTCTCTCTCTCTC	TCTCTCTCTCTCTC	TCTCTCTCTCTCTC	TCTCTCTCTCTCTC	TGCAG-ATCATCGAC	175
4 2A	TTCACTGAAGAGTCA	GGATGCCAAGACGGA	GTGAAGGACCTGAA	GACAATGCTACACAG	CAGGATGCAGATGTA	TGCTTAGGAGCGAG					155

Page 3.1

181	195	196	210	211	225	226	240	241	255	256	270
1 3B	TTT-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	
2 10B	TTCACTGAAGAGTCA	GGAT---	GCCAAGAC	GGAACTCGCTGCAGA	TCATCGACTTC						
3 4B	TTC-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	
4 2A	TTCGAAAGCCCTGGA	AACTTTAGAAAAAGGC	CATCGTCGCTGCAGA	TCATCGACTTC							

(26)

1 scores better than 57 saved, ktup: 4

```
initn initl opt
```

```
>NBISADFO Positive control F5/R5 NBISA #1 10/17 & 10/9 760 760 760
94.3% identity in 211 nt overlap
```

```

              70              80              90              100             110             120
2A DFO  ATGATTTCACCAGACATGGCGGAGGAGAAGACGATGTTGAAGGACCTGAAGACAATGCTA
          :::::::::: :::::::::: :::::::::: :::::::::: :::::::::: ::::::::::
NBISAD  ATGATTTCACCCGACATGGCAGAGGAGAAGACGATGCTGAAGGAGCTGAAAACAATGCTA
              70              80              90              100             110             120

```

2A DFO CACAGCAGGATGCAGATGTATGCTCTAGGAGCGAGTTCGAAAGCCCTGGAAACTTTAGAA
::: :
NBISAD CACAGCAGGATGCAGATGTATGCTCTGGGTGCAAGTTCGAAAGCCCTAGAGAATTTAGAA
130 140 150 160 170 180

```

                190          200          210
2A DFO AAGGCCATCGTCGCTGCAGATCATCGACTTC
      : ::::::::::::::::::::X
NBISAD AAGGCCATCGTCGCTGCAGATCATCGACTTC
                190          200          210

```

13

```
116 residues in      1 sequences
1 scores better than 57 saved, ktup: 4
```

```

10B DF CGACTTC
      :::::X
3B      CGACTTCGCTGCAGATCATCGACTTC
          100      110

```

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Jones, Simon

From: Dr. Fred Kibenge [kibenge@upei.ca]
Sent: December 8, 2003 6:59 AM
To: KibengeM@pac.dfo-mpo.gc.ca
Cc: JonesS@pac.dfo-mpo.gc.ca
Subject: ACGT information

Follow Up Flag: Follow up
Flag Status: Red

Hi Molly,

Here is the ACGT information you wanted:

They sequence PCR products.

Sequencing can be done with PCR primers used to generate the PCR product, but you would miss the primer sequence (i.e., the M13 Forward primer sequence or the M13 Reverse primer sequence would not show). If you want complete sequence then you have to ask to sequence with both primers separately.

In 10% of the sequencing reactions (that do not work) they may want to try internal primers, i.e., your F5 or R5 primers; but they would let you know if this is necessary.

To sequence a 500bp product, they need 50 ng per reaction. It is recommended that you pass your PCR product through the QIAGEN PCR purification kit, and then send them the purified material as is. They prefer that you also send them a photo of the gel (together with the sample for sequencing) indicating the volume of the PCR product that was loaded.

They charge \$30 per sequencing reaction. For a 500bp fragment, you need only one reaction.

Here is the contact information:
Ask for Chi or Philip.

ACGT Corporation
100 Bay Street, 11th Floor
Toronto, Ontario
M5G 1Z6

Tel: 1-800-735-0847 or
1-416-977-2228.

Fax: 1-416-977-7122.

You already have their e-mail address: order@acgtcorp.com.

Let me know if you need more information.

Fred.